

# Package ‘contrast’

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**Title** A Collection of Contrast Methods

**Version** 0.24.2

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**Description** One degree of freedom contrasts for 'lm', 'glm', 'gls', and 'geese' objects.

**License** GPL-2

**URL** <https://github.com/Alanocallaghan/contrast>

**BugReports** <https://github.com/Alanocallaghan/contrast/issues>

**Encoding** UTF-8

**LazyData** true

**Depends** R (>= 2.10)

**Imports** nlme, sandwich, rms

**Suggests** knitr, kableExtra, dplyr, ggplot2, tidyr, rmarkdown,  
testthat, covr, geopack, splines

**RoxygenNote** 7.2.1

**VignetteBuilder** knitr

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**NeedsCompilation** no

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contrast.lm	<i>General Contrasts of Regression Coefficients</i>
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### Description

This function computes one or more contrasts of the estimated regression coefficients in a fit from one of the functions in Design, along with standard errors, confidence limits, t or Z statistics, P-values.

### Usage

```
## S3 method for class 'lm'
contrast(fit, ...)

## S3 method for class 'gls'
contrast(fit, ...)

## S3 method for class 'lme'
contrast(fit, ...)

## S3 method for class 'geese'
contrast(fit, ...)

contrast_calc(
  fit,
  a,
  b,
  cnames = NULL,
  type = c("individual", "average"),
  weights = "equal",
  conf.int = 0.95,
  fcType = "simple",
  fcFunc = I,
  covType = NULL,
  ...,
  env = parent.frame(2)
)
```

**Arguments**

<code>fit</code>	A fit of class <code>lm</code> , <code>glm</code> , etc.
<code>...</code>	For <code>contrast()</code> , these pass arguments to <code>contrast_calc()</code> . For <code>contrast_calc()</code> , they are not used.
<code>a, b</code>	Lists containing conditions for all predictors in the model that will be contrasted to form the hypothesis $H_0: a = b$ . The <code>gendata</code> function will generate the necessary combinations and default values for unspecified predictors. See examples below.
<code>cnames</code>	A vector of character strings naming the contrasts when <code>type = "individual"</code> . Usually <code>cnames</code> is not necessary as the function tries to name the contrasts by examining which predictors are varying consistently in the two lists. <code>cnames</code> will be needed when you contrast "non-comparable" settings, e.g., you compare <code>list(treat = "drug", age = c(20, 30))</code> with <code>list(treat = "placebo", age = c(40, 50))</code> .
<code>type</code>	A character string. Set <code>type = "average"</code> to average the individual contrasts (e.g., to obtain a "Type II" or "Type III" contrast).
<code>weights</code>	A numeric vector, used when <code>type = "average"</code> , to obtain weighted contrasts.
<code>conf.int</code>	The confidence level for confidence intervals for the contrasts.
<code>fcType</code>	A character string: "simple", "log" or "signed".
<code>fcFunc</code>	A function to transform the numerator and denominator of fold changes.
<code>covType</code>	A string matching the method for estimating the covariance matrix. The default value produces the typical estimate. See <code>sandwich::vcovHC()</code> for options.
<code>env</code>	An environment in which evaluate <code>fit</code> .

**Details**

These functions mirror `rms::contrast.rms()` but have fewer options.

There are some between-package inconsistencies regarding degrees of freedom in some models. See the package vignette for more details.

Fold changes are calculated for each hypothesis. When `fcType = "simple"`, the ratio of the a group predictions over the b group predictions are used. When `fcType = "signed"`, the ratio is used if it is greater than 1; otherwise the negative inverse (e.g.,  $-1/\text{ratio}$ ) is returned.

**Value**

a list of class `contrast.Design` containing the elements `Contrast`, `SE`, `Z`, `var`, `df.residual`, `Lower`, `Upper`, `Pvalue`, `X`, `cnames`, and `foldChange`, which denote the contrast estimates, standard errors, Z or t-statistics, variance matrix, residual degrees of freedom (this is `NULL` if the model was not `ols`), lower and upper confidence limits, 2-sided P-value, design matrix, and contrast names (or `NULL`).

**See Also**

`rms::contrast.rms()`, `sandwich::vcovHC()`

**Examples**

```

library(nlme)
Orthodont2 <- Orthodont
Orthodont2$newAge <- Orthodont$age - 11
fm10rth.lme2 <- lme(distance ~ Sex * newAge,
  data = Orthodont2,
  random = ~ newAge | Subject
)
summary(fm10rth.lme2)

contrast(fm10rth.lme2,
  a = list(Sex = levels(Orthodont2$Sex), newAge = 8 - 11),
  b = list(Sex = levels(Orthodont2$Sex), newAge = 10 - 11)
)

# -----

anova_model <- lm(expression ~ diet * group, data = two_factor_crossed)
anova(anova_model)

library(ggplot2)
theme_set(theme_bw() + theme(legend.position = "top"))
ggplot(two_factor_crossed) +
  aes(x = diet, y = expression, col = group, shape = group) +
  geom_point() +
  geom_smooth(aes(group = group), method = lm, se = FALSE)

int_model <- lm(expression ~ diet * group, data = two_factor_crossed)
main_effects <- lm(expression ~ diet + group, data = two_factor_crossed)

# Interaction effect is probably real:
anova(main_effects, int_model)

# Test treatment in low fat diet:
veh_group <- list(diet = "low fat", group = "vehicle")
trt_group <- list(diet = "low fat", group = "treatment")
contrast(int_model, veh_group, trt_group)

# -----

car_mod <- lm(mpg ~ am + wt, data = mtcars)
print(summary(car_mod), digits = 5)

mean_wt <- mean(mtcars$wt)

manual_trans <- list(am = 0, wt = mean_wt)
auto_trans <- list(am = 1, wt = mean_wt)
print(contrast(car_mod, manual_trans, auto_trans), digits = 5)

```

---

```
print.contrast      Print a Contrast Object
```

---

**Description**

Print a Contrast Object

**Usage**

```
## S3 method for class 'contrast'
print(x, X = FALSE, fun = function(u) u, ...)
```

**Arguments**

x	Result of contrast().
X	A logical: set TRUE to print design matrix used in computing the contrasts (or the average contrast).
fun	A function to transform the contrast, SE, and lower and upper confidence limits before printing. For example, specify fun = exp to anti-log them for logistic models.
...	Not used.

---

```
two_factor_crossed  Complete Two-Factor Experiment
```

---

**Description**

Complete Two-Factor Experiment

**Details**

A gene expression experiment was run to assess the effect of a compound under two different diets: high fat and low fat. The main comparisons of interest are the difference between the treated and untreated groups within a diet. The interaction effect was a secondary hypothesis. For illustration, we only include the expression value of one of the genes. The study design was a full two-way factorial with n = 24 samples.

**Value**

```
two_factor_crossed
      A data frame
```

**Examples**

```
two_factor_crossed
```

---

two_factor_incompl	<i>Incomplete Two-Factor Experiment with Repeated Measurements</i>
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**Description**

Incomplete Two-Factor Experiment with Repeated Measurements

**Details**

In a gene expression experiment, stem cells were differentiated using a set of factors (such as media types, cell spreads etc.). These factors were collapsed into a single cell environment configurations variable. The cell lines were assayed over three days. Two of the configurations were only run on the first day and the other two were assayed at baseline.

To get the materials, three donors provided materials. These donors provided (almost) equal replication across the two experimental factors (day and configuration).

One of the goals of this experiment was to assess pre-specified differences in the configuration at each time point. For example, the differences between configurations A and B at day one is of interest. Also, the differences between configurations C and D at each time point were important.

Since there are missing cells in the design, it is not a complete two-way factorial. One way to analyze this experiment is to further collapse the time and configuration data into a single variable and then specify each comparison using this factor.

**Value**

two\_factor\_incompl  
A data frame

**Examples**

two\_factor\_incompl

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